Review paper

Prostaglandins in the treatment of cancer

Hiroshi Sasaki¹ and Mananori Fukushima²

¹Department of Obstetrics and Gynecology, The Jikei University School of Medicine, Minato-ku, Tokyo 105, Japan. Tel: (+81) 3 3433 1111, extn 3521; Fax: (+81) 3 5473 8421. ²Department of Internal Medicine, Aichi Cancer center, Chikusa-ku, Nagoya 464, Japan.

Prostaglandins (PGs) with antiproliferative activity against tumor cells consist of the cyclopentenone PGs and the alkylidene cyclopentenone PGs. Such PGs are PGD₂, PGJ₂, Δ^{12} -PGJ₂, PGA₁, Δ^{7} -PGA₁ and PGA₂. Both PGJ_2 and Δ^{12} - PGJ_2 are ultimate metabolites of PGD_2 and have potent antiproliferative activity on tumor cells. Δ^{12} - PGJ_2 was identified in human urine, whereas Δ^7 -PGA₁ has not been found in the human body. One important characteristic of both Δ^7 -PGA₁ and Δ^{12} -PGJ₂ is that they have little cross resistance with cisplatin and adriamycin in vitro and in vivo. Δ^7 -PGA₁ has 5-fold greater antitumor activity than $\Delta^{12}\text{-PGJ}_2$. Methyl ester- $\Delta^7\text{PGA}_1$ (methyl- Δ^7 -PGA₁) is stable chemically and can be easily synthesized In large amounts. All four isomers of methyl- Δ^7 -PGA₁ showed the same antiproliferative activities on ovarian carcinoma cells. In addition, methyl- Δ^7 -PGA₁ integrated in lipid microspheres (lipo-methyl- Δ^7 -PGA₁) is more soluble in water than methyl-Δ7-PGA₁ alone. Hence, lipomethyl- Δ^7 -PGA₁ was selected for extensive preclinical studies. Intravenous administration of lipo-methyl- Δ^7 -PGA₁ could inhibit the growth of both HeLa S3 and Lovo colon cancer cells transplanted subcutaneously in nude mice. Lipo-methyl- Δ^7 -PGA₁ by intraperitoneal administration could prolong the survival of scid mice bearing 2008C/13* cells resistant to cisplatin. The combined administration of cisplatin and lipo-methyl- Δ^7 -PGA₁ prolonged the survival of nude mice bearing HRA cells compared with each single agent alone. Consequently, phase 1 clinical studies of lipo-methyl- Δ^7 -PGA₁ in refractory ovarian cancer and colon cancer are planned. Δ^7 -PGA₁ demonstrates irreversible binding to thiols, whereas PGA₁ shows reversible binding. Both Δ^7 -PGA₁ and lipo-methyl- Δ^7 -PGA₁ are metabolized to unknown products in human serum, whereas the latter is converted to Δ^7 -PGA₁ in rat serum. The half-life of Δ^7 -PGA₁ was 1.5 h in human serum, whereas that of methyl- Δ^7 -PGA₁ was 13 min. The half-life of lipo-methyl- Δ^7 -PGA₁ was almost the same as that of methyl- Δ^7 -PGA₁. On the other hand, Δ^{12} -PGJ₂ was stable in human serum. Lipomethyl- Δ^7 -PGA₁ showed less toxicity than Δ^7 -PGA₁ and Δ^{12} -PGJ₂. LD₁₀ and LD₅₀ for single administration were 17.4 and 33.5 mg/kg for male rats, and 38.1 and 45.9 mg/kg for female rats, respectively. All rats tolerated repeated administration of 10 mg/kg/day lipo-methyl- Δ^7 -PGA₁ for 28 days. However, loss of weight was observed after 14 days of administration. Slight anemia

was recognized with decrease of both red blood cell count and hemoglobin. However, the dose-limiting factors remain undetermined. The inhibition of DNA synthesis by antitumor PGs is independent of AMP. PGs were transferred into the nucleus, and Δ^{12} -PGJ₂ covalently bound to nuclear proteins and inhibited RNA synthesis. With respect to their antiproliferative activity, the primary effect of PGA₁, PGD₂, Δ^7 -PGA₁ and Δ^{12} -PGJ₂ was to block cell progression from G_1 to S in the cell cycle. Both PGA₁ and PGJ₂ Induced the synthesis of a 70 kDa protein (p70) which was identified as a heat shock protein related to the major 70 kDa heat shock protein group. The G, block is associated with both inhibition of myc gene family expression and induction of heat shock proteins. High dose administration of either PGA₂ or Δ^{12} -PGJ₂ resulted in significant G2/M arrest and apoptosis. The PGs with a cyclopentenone ring have antiviral activity. PGJ₂ is a potent antiviral agent against Sendal virus and herpes simplex virus. In addition, PGA₁ and 16,16-dimethyl-PGA2-methylester suppressed the proliferation of HIV-1-infected cord blood lymphocytes in vitro. The potency of antiviral activity was Δ^7 -PGA₁ > PGA₂. The cyclopentenone ring seems to have a universal action of both antitumor activity and antiviral DNA activity.

Key words: Antitumor activity, antiviral activity, mechanisms, prostaglandins.

Introduction

Prostaglandins (PGs) have been shown to be involved in the regulation of many physiological phenomena, including cell growth and differentiation¹⁻⁵ and immune function,⁶⁻¹² and to possess antiviral activity. ¹³⁻¹⁸ In several recent studies, prostaglandins have shown antitumor activity at the level of cell differentiation and proliferation, and enhance the host's immunity against tumors. ⁶⁻¹² Most of the early studies on the antitumor effects of PGs examined the inhibition of animal tumors by PGA and PGE or their analogs. These PGs were shown to inhibit the growth *in vitro* and *in vivo* of several murine melanomas and Lewis lung carcinoma. Of the prostaglandins, PGD₂ has greater

potency and few side effects. $^{5,19-21}$ Identification of PGJ₂ was epoch making in the development of antitumor PGs, since it was found to exhibit more potent tumor suppression than PGD₂ without other PGD₂ activity. 22 The conversion of PGD₂ to Δ^{12} -PGJ₂ in the presence of serum or albumin was demonstrated. 23 Δ^{12} -PGJ₂ has the most potent antitumor activity of the derivatives of PGD₂. 22,23

 Δ^{12} -PGJ₂ was found in human urine and was proven to be produced in the human body.²⁴ This suggests that Δ^{12} -PGJ₂ has some physiological roles, such as inhibition of tumors or viruses. PGA derivatives also have in vivo antitumor activity. Recently, both Δ^7 -PGA₁ and PGJ₂ have been continued to be investigated for antitumor²⁵⁻³⁰ and antiviral activities.31-33 However, their precise mechanism of action is still unclear. Preclinical studies have continued to identify target tumors of both Δ^7 - PGA_1 and Δ^{12} - PGJ_2 , particularly in relation to drug-resistant tumors. ^{27,28} This supports the clinical application of antitumor PGs for drug-resistant tumors. At present, one such antitumor PG, lipomethyl- Δ^7 -PGA₁, has been selected for clinical studies, and both its antitumor activity in vivo and toxicity profile have been investigated. Hence, the present review focuses on the preclinical studies and potential of PGs as anticancer drugs.

Chemical structure of antitumor PGs

PGs that have antiproliferative effects on tumor cells are PGD₂, PGJ₂ Δ^{12} -PGJ₂, PGA1, Δ^{7} -PGA1 and PGA₂. Both PGJ₂ and Δ^{12} -PGJ₂ are ultimate metabolites of PGD₂,²³ and have potent antiproliferative activity on tumor cells. The chemical structures of the PGs responsible for their antiproliferative activity are cyclopentenone and alkylidene cyclopentenone.34 Alkylidene cyclopentenone PGs such as Δ^{12} -PGJ₂ and Δ^{7} -PGA₁ have the most potent antiproliferative activity on tumor cells of the cyclopentenone PGs. Some octocorals produce Δ^7 -PGA type alkylidene cyclopentenone eicosanoids such as clavulone and punaglandin which have similar antiproliferative activity to Δ^{12} -PGJ₂ and Δ^{7} -PGA₁. Such PGs contain 2-enone structures and are highly reactive with nucleophilic agents, thus forming Michael adducts.34-36 This chemical reactivity may explain the biochemical activity of alkylidene cyclopentenone eicosanoid PGs. Modification of the Δ^7 PGA₁ α - or β -side chains was performed to study the relation between the hydrophilicity/lipophilicity and the antiproliferative activity in vitro. Lipophilic derivatives of Δ^7 -PGA₁ possess more potent anti-

proliferative activity on cultured cells in vitro than hydrophobic derivatives.³⁷ Δ^7 -PGA₁ and the Δ^7 -PGA₁ analog 5-[(E)-4,7-dihydroxy-(2E)-pentenylidene]-4-hydroxy-4-(4-phenoxybutyl)-2-cyclopentenone (TE10303) showed potent antiproliferative activity in ovarian cancer in vitro and in vivo*.38 Drug penetration of TE10303 into a multicellular spheroid was to a depth of about 150 µm from the outer layer. This suggests that hydrophobic PGs have poor tumor penetration (Sasaki and Terashima, unpublished observation). The optimal chemical structure of PGs for intraperitoneal or intravenous administration may be different from that of the potent antiproliferative PGs. The chemical stability of alkylidene cyclopentenone eicosanoids has been investigated in relation to their reaction with SH-containing compounds. Glutathione (GSH) reacts with Δ^7 -PGA₁ methyl ester with a larger dissociation constant at neutral pH than with PGA₁ methyl ester. 36 Δ^7 -PGA₁ demonstrates irreversible binding. Δ^{12} -PGJ₂ was identified in human urine, but Δ^7 -PGA₁ has not been found in the human body. Methyl- Δ^7 -PGA₁ is one of the synthesized PGs. Δ^7 -PGA₁³⁸ and methyl- Δ^7 -PGA¹ (Sasaki and Terashima, unpublished observation) have shown antiproliferative activity in vivo. Methyl- Δ^7 -PGA₁ exhibits potent inhibition of cell growth in vitro; 26 however, it showed weak effects in vivo. 39 This suggests that methyl- Δ^7 -PGA₁ may be metabolized in the human body. Furthermore, investigations are necessary on the metabolism of Δ^7 -PGA₁ analogs in the human body. Four isomers of methyl- Δ^7 -PGA₁ were synthesized and investigated for their antiproliferative activity on human ovarian carcinoma cells (2008 and A2780 cell lines). All isomers of methyl- Δ^7 -PGA₁ showed the same antiproliferative activities on ovarian carcinoma cells. 40 This suggests that there may be no stereospecific receptors for the antiproliferative effects of methyl- Δ^7 -PGA₁. Methyl- Δ^7 -PGA₁ is chemically stable and can be easily synthesized in large amounts. In addition, methyl- Δ^7 -PGA₁ integrated into lipid microspheres (lipo-methyl- Δ^7 -PGA₁) is more soluble in water than methyl- Δ^7 -PGA₁ alone. Hence, lipo-methyl- Δ^7 -PGA₁ was selected for preclinical studies.³⁹

Antitumor activity

Antiproliferative activity in vitro

The combination of PGE₂ and tumor necrosis factor (TNF) was effective in inhibiting proliferation of U-

937, ML-1 and HL-60 cells. 41 The effective concentration of PGE₂ was 100 ng/ml. This concentration of PGE₂ could induce the differentiation of U-937 cells. PGD_2 , PGJ_2 and Δ^{12} - PGJ_2 either alone or in combination with TNF at a concentration of 100 ng/ ml did not induce any differentiation in U-937 cells. However, the combination of TNF and Δ^{12} -PGJ₂ could inhibit the proliferation of three cultured cell lines (HeLa S3, HHUA and CAOV-3) derived from gynecologic malignancies. 42 The effective concentration of Δ^{12} -PGJ₂ was 1 µg/ml, which was less than that of PGE2. Synergism between a series of PGs and interferon (IFN)-α has also been reported. 43 Idomethacin, an inhibitor of PGs, could modify tumor growth. 44,45 This could be derived from synergism between PGs and interleukin, IFNs or growth factors. The interaction between PGs and cytokines may play a role in the antitumor activity of PGs.

The antiproliferative effects of PGs alone on tumor cells continue to be studied for PGD₂ and the cyclopentenone PGs. ^{34,46} PGD₂ inhibited growth in over 20 different cancer cell lines. 47-49 The spectrum of antiproliferative activity of PGD₂ against tumor cells in vitro demonstrated that PGD2 was active against human osteosarcoma, breast cancer, rhabdomyosarcoma, uterine cervical carcinoma, ovarian cancer and melanoma cells at a concentration of 5 µg/ml.⁴⁸ The antiproliferative activity of PGD2 can be interpreted as being mediated by Δ^{12} -PGJ₂ which is the ultimate metabolite of PGD₂. PGD₂ is converted to Δ^{12} -PGJ₂ by human albumin and serum. 23,50 Δ^{12} -PGJ₂ has 5-fold greater antiproliferative activity on L1210 cells than PGD₂.²³ Kato et al. demonstrated antitumor activity of Δ^7 - PGA_1 and Δ^{12} - PGJ_2 in vitro and in vivo. 51 Ikai et al. showed that Δ^{12} -PGJ₂ destroyed the cytoskeleton of transformed epidermal cells in culture on the basis of inhibition of protein synthesis. 52 The authors reported that $\Delta^{12}\text{-PGJ}_2$ and an analog of PGA1 could inhibit the growth of ovarian dysgerminoma cells in vitro. 38 Kikuchi showed that the cell proliferation of HRA cells was inhibited by Δ^{12} -PGJ₂ and an analog of PGA₁ in vitro.²⁹ Mitsuoka et al. reported an inhibitory effect of prostaglandin Δ^{12} - PGJ_2 on proliferation and α -fetoprotein expression in HuH-7 human hepatoma cells. ²⁵ Methyl- Δ^7 -PGA₁ and a Δ^7 -PGA₁ analog (TE10303) based on modification of the side chains were synthesized and were shown to have potent inhibitory activity on cell proliferation in vitro. 37,38,50 Δ^7 -PGA₁ has 5-fold greater antitumor activity than Δ^{12} -PGJ₂. One particularly important characteristic of both Δ^7 -PGA₁ and Δ^{12} -PGJ₂ from the clinical aspect is their low cross resistance both to cisplatin and to adriamycin.

The authors studied the antitumor activity of both Δ^7 -PGA₁ and Δ^{12} -PGJ₂ on human ovarian cancer cell lines resistant to cisplatin, doxorubicin (ADR) and L-phenylalanine mustard (L-PAM). Drug resistance ratios based on IC50 were 62.5 for ADR in A2780^{AD} cells⁵³ and 16.0 for cisplatin in 2008DDP cells, ⁵⁴ whereas the ratios for Δ^7 -PGA₁ and Δ^{12} -PGJ₂ were 1.5 and 1.8 in A2780^{AD} cells, and 2.3 and 3.2 in 2008DDP cells, respectively.²⁸ A2780^{AD} cells express the mdr gene, which is a major factor in their drug resistance.⁵⁵ DNA repair, a high level of intracellular glutathione and high activity of protein kinase C are thought to be factors related to drug resistance in 2008DDP cells.⁵⁴ It is an interesting phenomenon that both Δ^7 -PGA₁ and Δ^{12} -PGJ₂ can overcome the factors of drug resistance.

The antiproliferative activity of PGs has been thought to be mediated by AMP. 4,55 However, Wiley et al. first reported that antitumor PG-induced inhibition of DNA synthesis is independent of AMP, because dideoxyadenosine, which is an inhibitor of adenylate cyclase, could not inhibit DNA synthesis.⁵⁶ In addition, Hughes-Fulford showed that PGJ2 did not affect the intracellular level of AMP, while it inhibited DNA synthesis.⁵⁷ Narumiya and Fukushima measured the intracellular level of AMP of cancer cells treated with PGE₂, BW245C, PGD₂ and Δ^{12} -PGJ₂. They showed that Δ^{12} -PGJ₂ never increased the cellular cAMP level.⁵⁸ Analyses of the intracellular uptake and distribution of $\Delta^{12}\text{-PGJ}_2$ and PGA_2 demonstrated that intracellular uptake occurred above 20°C and that both PGs were transferred into the nucleus at 37°C.59 However, Δ^{12} -PGJ₂ differs from PGA₂ in relation to intracellular pharmacokinetics. Intracellular PGA2 was washed out with PG-free medium and less than 15% of the initial amount remained in the nuclei, whereas 70% of the initial amount of Δ^{12} -PGJ₂ remained in the nuclei after intensive washing. Both hypotonic washing and 1% Triton X-100 solvent could not release Δ^{12} -PGJ₂ from the nuclei. Chromatographic analysis demonstrated that Δ^{12} -PGJ₂ was covalently bound to nuclear proteins.⁶⁰ These findings suggest that Δ^{12} -PGJ₂ binding in the nucleus was more stable than that of other PGs.

Bhuyan *et al.* reported that the primary effect of PGA_1 , PGA_2 and PGD_2 was to block cell progression from G_1 to S in the cell cycle. At 2.5 μ g/ml of PGD_2 , blockade of cells in G_1 and normal progression through the other phases resulted in accumulation of 80–90% of the cells in G_1 . At this concentration, there was no inhibition of DNA synthesis, and cells in S progressed apparently normally through S until

all cells were blocked in G₁.61 Marui et al. demonstrated that the G1 block was induced by both inhibition of N-myc gene expression and induction of heat shock proteins.⁶² In addition, Holbrook et al. measured mRNA of the HSP70 gene family and showed that high-level induction is specific to HSP70 mRNA, resulting in an increase in the rate of transcription. Cycloheximide pretreatment, which inhibits the antiproliferative effects of PGA₂, prevents activation of the heat shock factor and induction of HSP70 mRNA by PGA2.63 In relation to the cell cycle modifying effects of PGs, G₂/M arrest has been reported. Treatment of L1210 cells with PGA₂ or Δ^{12} -PGJ₂ resulted in significant G₂/M arrest and subsequent DNA fragmentation at concentrations that are cytotoxic to the cells. When the morphology of cells was examined by electron microscopy, L1210 cells incubated with a cytotoxic dose of PGA2 or Δ^{12} -PGJ2 for 24 h showed characteristic morphological features of apoptosis such as chromatin condensation, nuclear fragmentation and formation of apoptotic bodies.⁶⁴ However, since this is only one report, further investigations are necessary regarding the induction of apoptosis by PGs.

Antiproliferative activity in vivo

Since the 1970s it has been well known that PGs of the E and A series inhibit the growth of some lines of tumor cells in vivo. 65,66 PGD₂ was also found to be a potent antitumor agent in vivo. 51,67 However, the discovery of PGJ2 led to further studies of antitumor PGs including the PGA series, PGJ series and punaglandin. The antitumor activity of PGs in vivo is mainly classed as two types. One is derived from the immune effects of PGs. Combined administration of PGD₂ and cisplatin exhibited additive effects on human ovarian cancer cell growth in nude mice. 11,19,20,68,69 When antineoplastic PGs were administered to nude mice bearing human ovarian cancer cells, tumor growth in groups treated with PGJ_2 and Δ^{12} - PGJ_2 alone was significantly inhibited 63 days after tumor inoculation, compared to that in an untreated group.⁷⁰ Combination of Δ^7 -PGA₁ and cisplatin could prolong survival of nude mice bearing human ovarian cancer cells.²⁹ Kita et al. demonstrated that Δ^{12} -PGJ₂ dose-dependently stimulated phytohemagglutinin (PHA)-induced blast formation of human lymphocytes in the concentration range of 10^{-8} to 10^{-6} M.^{10,71} This may imply that the antitumor activity of antitumor PGs is based in part on their immune effects.

The other type of antitumor effects of PGs is derived from direct inhibition of tumor growth. The potency of antiproliferative activities of PGs on cultured cells originated from cancer is punaglandin > Δ^7 -PGA₁ > Δ^{12} -PGJ₂. ⁷⁰ However, punaglandin failed to produce significant prolongation of survival in vivo, while it induced significant weight loss. 70 Therefore, derivatives of Δ^7 -PGA₁ were synthesized and their antitumor activities were tested in in vivo models. Although lipophilic analogs of Δ^7 -PGA₁ have potent antitumor activity in vitro, hydrophobic analogs have greater activity than lipophilic analogs in vivo.37 TE10303, which was obtained by modification of the side chains, showed some local effects of human ovarian dysgerminoma transplanted in nude mice.³⁸ However, TE10303 could not inhibit the growth of tumor transplanted subcutaneously in nude mice on intraperitoneal administration. Lipo-methyl- Δ^7 -PGA₁ has been tested as a preclinical agent *in vivo*. ⁴⁰ It has been shown to inhibit the growth of both HeLa S3 and Lovo colon cancer cells transplanted subcutaneously in nude mice when administered intravenously.72 Lipomethyl- Δ^7 -PGA₁ by intraperitoneal administration could prolong the survival of scid mice bearing 2008C/13* cells resistant to cisplatin.54 The combination of cisplatin and lipo-methyl- Δ^7 -PGA₁ prolonged survival of nude mice bearing HRA cells compared to each single agent alone.⁷² These findings suggest that lipo-methyl- Δ^7 -PGA₁ could be a candidate for phase I and II clinical tudies to test its activity against refractory ovarian cancer and colon cancer.

Antiviral activity

It is well known that some viruses can cause cancer. Human papilloma virus and herpes simplex virus are thought to be causes of cancer of the uterine cervix. HTLV-1 virus causes T cell leukemia. Recently, AIDS associated cancers, including Kaposi sarcoma, lymphoma and cervical cancer, have been reported. AIDS Antitumor prostaglandins that can inhibit viral replication may be useful for the treatment of virus-associated cancers.

PGs of the A and J series inhibit viral replication. 34,46 In 1980, Santoro *et al.* reported antiviral effects of PGA compounds and furthermore demonstrated that PGJ₂ was a potent antiviral agent against Sendai virus. B D'Onofrio *et al.* demonstrated that PGA₁ and 16,16-dimethyl-PGA₂-methylester (diM-PGA₂) blocked the proliferation of HIV-1-infected cord blood lymphocytes (CBL) *in*

vitro. 76 Yamamoto et al. investigated the relation between chemical structure and antiviral activity using herpes simplex virus. They showed that cyclopentenone PGs have potent antiviral activity. The potency of antiviral activity was Δ^7 -PGA₁ > PGA₁ > PGA₂ and Δ^{12} -PGJ₂ > PGJ₂ > PGD₂.⁷⁷ The mechanisms of antiviral activities proposed are (i) inhibition of primary transcription of the viral genome, ⁷⁷ (ii) alterations in the synthesis and/or maturation of specific viral proteins, 18,78 (iii) inhibition of mutant mRNA synthesis at 39°C⁷⁹ and (iv) inhibition of transcription of viral genomic RNA.⁷⁹ PGA and PGJ have been shown to affect RNA replication from DNA in the nucleus and, thus, inhibit viral proliferation. The cyclopentenone ring seems to have a universal action of both antitumor activity and antiviral DNA activity.

Toxicity

The antitumor PG lipo-methyl- Δ^7 -PGA₁ is a candidate for clinical study. 39,46 Therefore, both the pharmacokinetics and toxicity of lipo-methyl- Δ^7 -PGA₁ in rats have been investigated. 72 Both Δ^7 -PGA₁ and lipo-methyl- Δ^7 -PGA₁ are metabolized to unknown compounds in human serum; however, it is converted to Δ^7 -PGA₁ in rat serum. The half-life of Δ^7 -PGA₁ was 1.5 h in human serum and that of methyl- Δ^7 -PGA₁ was 13 min. The half-life of lipomethyl- Δ^7 -PGA₁ was almost the same as that of methyl- Δ^7 -PGA₁. On the other hand, Δ^{12} -PGJ₂ was stable in human serum.⁷² This suggests that lipomethyl- Δ^7 -PGA₁ has lower toxicity than Δ^7 -PGA₁ and Δ^{12} -PGJ₂. When lipo-methyl- Δ^{7} -PGA₁ was intravenously administered to rats once [Sprague-Dawley rats, cri:CD (SD), 4 weeks after birth], LD_{10} and LD_{50} were 17.4 and 33.5 mg/kg for male rats, and 38.1 and 45.9 mg/kg for female rats, respectively. All rats tolerated repeated administration of 10 mg/kg/day lipo-methyl- Δ^7 -PGA₁ for 28 days. However, weight loss was observed after 14 days of administration. The dosage of 2.5 mg/kg/day did not induce loss of weight. Fifty percent of rats administered 10 mg/kg/day lipo-methyl- Δ^7 -PGA₁ exhibited both reduced movement and increased respiratory rate from 3 min after administration, which showed recovery by 20 min after administration. Appetite loss continued throughout the administration period.⁷²

In the urine examination, volume, color, SG, pH, protein, glucose, ketone bodies, bilirubin, urobilinogen, red blood cell count (RBC) and casts were within normal limits. Ocular examination was with-

in normal limits. Hematologic tests revealed anemia with decrease in both RBC and hemoglobin. However, both leukocytes and platelets showed no differences compared to control. Serum albumin concentration increased and A/G ratio increased. Adrenal gland weight decreased with administration of 5 mg/kg of lipo-methyl- Δ^7 -PGA₁ and higher doses. Furthermore, the relative weights of both lungs and heart to body weight increased with administration at 2.5 mg/kg/day and higher in rats. The weights of other organs did not change. Histopathological examination is now on going. These results suggest that the dose-limiting toxicity is effects on the adrenal glands, lungs and heart.

Conclusions

Cyclopentenone PGs and alkylidene cyclopentenone PGs have antiproliferative activity. The alkylidene cyclopentenone PGs Δ^{12} -PGJ₂ and Δ^{7} -PGA₁ have the most potent antiproliferative activity on tumor cells, and both exhibit antiviral and antitumor activity. Δ^{12} -PGJ₂ was identified in human urine, whereas Δ^7 -PGA₁ has not been found in the human body. Both Δ^7 -PGA₁ and Δ^{12} -PGJ₂ have little cross resistance both to cisplatin and to ADR. Δ^7 -PGA₁ has 5-fold greater antitumor activity than Δ^{12} - PGJ_2 . Methyl ester- Δ^7 - PGA_1 is stable chemically and can be easily synthesized in large amounts. In addition, methyl- Δ^7 -PGA₁ integrated into microspheres (lipo-methyl- Δ^7 -PGA₁) is more soluble in water than methyl- Δ^7 -PGA₁ alone. Preclinical *in vivo* studies of lipo-methyl- Δ^7 -PGA₁ have been conducted. Lipo-methyl- Δ^7 -PGA₁ has been shown to inhibit growth of ovarian cancer cells resistant to cisplatin. In addition, the combination of cisplatin and lipo-methyl- Δ^7 -PGA₁ showed additive antitumor effects on ovarian cancer cells in vivo. Lipo-methyl- Δ^7 -PGA₁ showed low toxicity in rats. The toxic effects in rats consisted of decrease in RBC and hemoglobin, reduction of adrenal gland weight, and increase of relative weight of lungs and heart. This suggests that lipo-methyl- Δ^7 -PGA₁ may be a candidate for phase I and II clinical studies to test its activity against refractory ovarian cancer and colon cancer.

Acknowledgements

This work was supported by a grant-in-aid, 'A comprehensive 10-year strategy for cancer control',

from the Ministry of Health and Welfare of Japan (1990–1993). The authors thank Dr Wendy Gray for her help in the preparation of the manuscript.

References

- Santoro MG, Philpott GW, Jaffe BM. Inhibition of B16 melanoma growth in vivo by a synthetic analog of prostaglandin E₂. Cancer Res 1977; 37: 3774-9.
- Bregman MD, Sander D, Meyskens FL Jr. Prostaglandins A₁ and E₁ inhibit the plating efficiency and proliferation of murine melanoma cells (Cloudman S-91) in soft agar. Biochem Biophys Res Commun 1982; 104: 1080-6.
- Honn KV, Romine M, Skoff A. Prostaglandin analogs as inhibitors of tumor cell DNA synthesis. Proc Soc Exp Biol Med 1981; 166: 562-7.
- Honn KV, Bockman RS, Marnett LJ. Prostaglandins and cancer: a review of tumor initiation through tumor metastasis. *Prostaglandins* 1981; 21: 833-64.
- Fukushima M, Kato T, Ueda R, Ota K, et al. Prostaglandin D2, a potential antineoplastic agent. Biochem Biophys Res Commun 1982; 105: 956-64.
- Fitzpatrick FA, Stringfellow DA. Prostaglandin D₂ formation by malignant melanoma cells correlates inversely with cellular metastatic potential. *Proc Natl Acad Sci USA* 1979; 76: 1765-9
- Stringfellow DA, Fitzpatrick FA. Prostaglandin D₂ controls pulmonary metastasis of malignant melanoma cells. Nature 1979; 282: 76–8.
- 8. Sanford MG, Philpott GD, Jaffe BM. Inhibition of tumour growth *in vivo* and *in vitro* by prostaglandin E. *Nature* 1976; **263**: 777-9.
- 9. Tsunamoto K, Todo S, Imashuku S. Effects of 5-bromo-2'-deoxyuridine on arachidonic acid metabolism of neuroblastoma and leukemia cells in culture: a possible role of endogenous prostaglandins in tumor cell proliferation and differentiation. *Prostaglandins Leukotrienes Med* 1987; 26: 157–69.
- 10. Honn KV, Cicone B, Skoff A. Prostacyclin: a potent antimetastatic agent. Science 1981; 212: 1270-2.
- 11. Kikuchi Y, Kita T, Hirata J, et al. Modulation of human lymphocyte response to phytohemagglutinin by antineoplastic prostaglandins. *Int J Immunopharmacol* 1992; **14**: 105–10.
- Keyaki A, Handa H, Yamashita J, et al. Growth-inhibitory effect of prostaglandin D₂ on mouse glioma cells. J Neurosurg 1984; 61: 912-7.
- Santoro MG Benedetto A, Carruba G, et al. Prostaglandin A compounds as antiviral agents. Science 1980; 209, 1032–4.
- 14. Santoro MG, Carruba G, Garaci E, et al. Prostaglandins of the A series inhibit Sendai virus replication in cultured cells. J Gen Virol 1981; 53: 75–83.
- Santoro MG, Jaffe BM, Elia G, et al. Prostaglandin A1 induces the synthesis of a new protein in cultured AGMK cells. Biochem Biophys Res Commun 1982; 107: 1179– 84.
- Benavente J, Esteban M, Jaffe BM, et al. Selective inhibition of viral gene expression as the mechanism of the antiviral action of PGA₁ in vaccinia virus-infected cells. J Gen Virol 1984; 65: 599–608.
- 17. Ankel H, Mittnacht S, Jacobsen H. Antiviral activity of

- prostaglandin A on encephalomyocarditis virus-infected cells: a unique effect unrelated to interferon. *J Gen Virol* 1985; **66**: 2355–64.
- Santoro MG, Fukushima M, Benedetto A, et al. PGJ₂, a new antiviral prostaglandin: inhibition of Sendai virus replication and alteration of virus protein synthesis. J Gen Virol 1987; 68: 1153–8.
- Kikuchi Y, Miyauchi M, Omori K, et al. Inhibition of human ovarian cancer cell growth in vitro and in nude mice by prostaglandin D₂. Cancer Res 1986; 46: 3364-6.
- Kikuchi Y, Miyauchi M, Iwano I, et al. Adjuvant effects of prostaglandin D₂ to cisplatin on human ovarian cancer cell growth in nude mice. Eur J Cancer Clin Oncol 1988; 24: 1829-33.
- Giles H, Leff P. The biology and pharmacology of PGD₂. Prostaglandins 1988; 35: 277-300.
- 22. Fukushima M, Kato T, Ota K, et al. 9-Deoxy- Δ^9 -prostaglandin D₂, a prostaglandin D₂ derivative with potent antineoplastic and weak smooth muscle-contracting activities. Biochem Biophys Res Commun 1982; 109: 626-33.
- Kikawa Y, Narumiya S, Fukushima M, et al. 9-Deoxy-Δ⁹, Δ¹²-13,14-dihydroprostaglandin D₂ a metabolite of prostaglandin D₂ formed in human plasma. Proc Natl Acad Sci USA 1984; 81: 1317-21.
- 24. Hurata Y, Hayashi H, Ito S, et al. Occurrence of 9-deoxy- Δ^9 , Δ^{12} -13,14-dihydroprostaglandin D^2 in human urine. Biol Chem 1988; **263**: 16619–25.
- 25. Mitsuoka S, Otsuru A, Nakao K, *et al.* Inhibitory effect of prostaglanding Δ^{12} -PGJ₂ on cell proliferation and α -fetoprotein expression in HuH-7; human hepatoma cells. *Prostaglandins* 1992; **43**: 189–97.
- Choi AMK, Fargnoli J, Carlson SG, et al. Cell growth inhibition by prostaglandin A₂ results in elevated expression of gadd153 mRNA. Exp Cell Res 1992: 199: 85-9.
- Amici C, Sistonen L, Santoro MG, et al. Antiproliferative prostaglandins activate heat shock transcription factor. Proc Natl Acad Sci USA 1992; 89 6227–31.
- 28. Sasaki H, Takada K, Terashima Y, et al. Human ovarian cancer cell lines resistant to cisplatin, doxorubicin, and L-phenylalanine mustard are sensitive to Δ^7 -prostaglandin A₁ and Δ^{12} -prostaglandin J Gynecol Oncol 1991; **41** 36–40.
- Kikuchi Y, Kita T, Miyauchi M, et al. Adjuvant effects of antineoplastic prostaglandins to cisplatin in nude mice bearing human ovarian cancer cells. J Cancer Res Clin Oncol 1992; 118: 453-7.
- 30. Shimakura S, Boland CR. Eicosanoid production by the human gastric cancer cell line AGS and its relation to cell growth. *Cancer Res* 1992; **52**: 1744–9.
- Sienko S, Eis-Hubinger AM, Schneweis KE. The role of free radical scavengers, inhibitors of prostaglandin synthesis, and hypomethylating agents in reactivation of latent herpes simplex virus. *Med Microbiol Immunol* 1991; 180: 249-59.
- 32. D'Onofrio C, Amici C, Puglianiello A, *et al.* Comparative anti-viral and anti-proliferative activity of PGA₁ and PGJ₂ against HTLV-1-infected MT-2 cells. *Int J Cancer* 1992; **51**: 481–8.
- Lum LSY, Hsu S, Vaewhongs M, et al. The hsp70 gene CCAAT-binding factor mediates transcriptional activation by the adenovirus E1a protein. Mol Cell Biol 1992; 12: 2599–605.
- 34. Fukushima M. Prostaglandin J₂ anti-tumour and anti-viral

- activities and the mechanisms involved. *Eicosanoids* 1990; **3**: 189–99.
- 35. Atsmon J, Sweetman BJ, Baertschi SW, et al. Formation of thiol conjugate of 9-deoxy- Δ^9 , Δ^{12} (E)-prostaglandin D₂ and Δ^{12} (E)-prostaglandin D₂. Biochemistry 1990; **29**: 3760-5.
- Noyori R, Suzuki M. Organic synthesis of prostaglandins: advancing biology. Science 1993: 259 44-5.
- Fukushima M, Suzumura Y, Kato T, et al. Antitumor prostaglandins (PGs): antitumor activity related to lipophilic character. Proc Am Ass Cancer Res 1988; 29: 329.
- 38. Sasaki H, Terashima Y, Takada K, et al. Antitumor effects of $12\text{-OH-}\Delta^{12}$ prostaglandin (PG) A_1 analogue on human ovarian cancer transplanted in nude mice. Proc Am Ass Cancer Res 1989; 30: 582.
- 39. Fukushima S, Takeuchi Y, Nakano M, et al. Δ^7 -PGA₁ lipid microspheres pharmaceutical properties and preclinical evaluations. In: Yasugi T, Nakamura H, Soma M, eds *Advances in polyunsaturated fatty acid res.* Tokyo: Excerpta Medica 1993; 67–70.
- Sasaki H, Terashima Y, Fukushima M, et al. Cell cycle arrest at G₁ phase by antitumor prostaglandins. The cell cycle '93: Regulators, targets and clinical applications. Proc XIIIth Washington International Spring Symp 1993; 56.
- Hosoi T, Takeda K, Konno K. Synergism of prostaglandin
 E₂ plus TNF in induction of differentiation of human monocytoid leukemic U-937 cells. Anticancer Res 1989; 9: 615-8.
- Mori H, Takada Y, Kondoh N, et al. Augmentation of antiproliferative activity of recombinant human tumor necrosis factor by Δ¹²-prostaglandin J₂. J Biol Response Mod 1990; 9: 260-3.
- 43. Bregman MD, Fukushima M. Modulation of melanoma by antineoplastic prostaglandins. In: Kimura K, Ota K, Herberman RB, Takita H, eds Cancer chemotherapy challenges for the future II. Tokyo: Excerpta Medica 1987: 82-90.
- 44. Gati I, Bergstrom M, Westerberg G, et al. Effects of prostaglandin and leukotriene inhibitors on the growth of human glioma spheroids. Eur J Cancer 1990; 26: 802-7.
- 45. Gati I, Bergstrom M, Csoka K, et al. Effects of the 5-lipoxygenase inhibitors AA-863 and U-60,257 on human glioma cell lines. *Prostaglandins Leukotrienes Essential Fatty Acids* 1990; 40: 117-24.
- Fukushima M. Biological activities and mechanism of action of PGJ₂ and related compounds: an update. Prostaglandins Leukotrienes Essential Fatty Acids 1992; 47: 1-12.
- Simmet T, Jaffe BM. Inhibition of B-16 melanoma growth in vitro by prostaglandin D₂. Prostaglandins 1983; 25: 47-54.
- Sakai T, Yamaguchi N, Shiroko Y, et al. Prostaglandin D₂ inhibits the proliferation of human malignant tumor cells. Prostaglandins 1984; 27: 17-26.
- Sakai T, Yamaguchi N, Kawai K, et al. Prostaglandin D₂ inhibits the proliferation of human neuroblastoma cells. Cancer Lett 1983; 17: 89-94.
- Fitzpatrick FA, Wynalda MA. Albumin-catalyzed metabolism of prostaglandin D₂. Identification of products formed in vitro: J Biol Chem 1983: 258 11713-8.
- 51. Kato T, Fukushima M, Kurozumi S, et al Antitumor activity of Δ^7 -prostaglandin A_1 and Δ^{12} -prostaglandin J_2 in vitro and in vivo. Cancer Res 1986; **46**: 3538-42.

- Ikai K, Fukushima M. Effects of cytotoxic prostaglandin, Δ¹²-PGJ₂ on protein synthesis and cytoskeleton in transformed epidermal cells in culture. Arch Dermatol Res 1990: 282: 131-4.
- Louie KG, Hamilton TC, Winker MA, et al. Adriamycin accumulation and metabolism in adriamycin-sensitive and -resistant human ovarian cancer cell lines. Biochem Pharmacol 1986: 35: 467-72.
- Andrews PA, Velury S, Mann SC, et al. Cis-diamminedichloroplatinum (II) accumulation in sensitive and resistant human ovarian carcinoma cells. Cancer Res 1988;
 48: 68-73.
- Jaffe BM, Santoro MG. Prostaglandins and cancer. In: Ramwell PW, ed., *The prostaglandins* New York: Plenum 1977: 329–51.
- Wiley MH, Fiengold KR, Grunfeld C, et al. Evidence for cAMP-independent inhibition of S-phase DNA synthesis by prostaglandins. J Biol Chem 1983; 258: 491-6.
- 57. Hughes-Fulford M, Wu J, Kato T, et al. Inhibition of DNA synthesis and cell cycle by prostaglandins independent of cyclic AMP. Adv Prostaglandin, Thromboxane, and Leukotriene Res 1985; 15: 401-4.
- Narumiya S, Fukushima M. Δ¹²-Prostaglandin J₂, an ultimate metabolite of prostaglandin D₂ exerting cell growth inhibition. *Biochem Biophys Res Commun* 1985: 127: 739–45.
- Narumiya S, Ohno K, Fujiwara M, et al. Site and mechanism of growth inhibition by prostaglandins. II. Temperature-dependent transfer of a cyclopentenone prostaglandin to nuclei. J Pharmacol Exp Ther 1986: 239: 506-11.
- 60. Narumiya S, Ohno K, Fukushima M, et al. Site and mechanism of growth inhibition by prostaglandins. III. Distribution and binding of prostaglandin A_2 and Δ^{12} -prostaglandin J_2 in nuclei. J Pharmacol Exp Ther 1987; 242: 306–11.
- 61. Bhuyan BK, Adams EG, Badiner GJ, et al. Cell cycle effects of prostaglandins A₁, A₂, and D₂ in human and murine melanoma cells in culture. Cancer Res 1986; **46**: 1688–93
- Marui N, Nishino H, Sakai T, et al. Δ¹²-Prostaglandin J₂ mimics heat shock in inducing cell cycle arrest at G₁ phase. Biochem Biophys Res Commun 1991; 179: 1662-9.
- 63. Holbrook NJ, Carlson SG, Choi AMK, et al. Induction of HSP70 gene expression by the antiproliferative prostaglandin PGA₂: a growth-dependent response mediated by activation of heat shock transcription factor. Mol Cell Biol 1992; 12: 1528-34.
- 64. Kim IK, Lee JH, Sohn HW, et al. Prostaglandin A₂ and Δ¹²-prostaglandin J₂ induce apoptosis in L1210 cells. FEBS Lett 1993; 321, 209-14.
- Eisenbarth GS, Wellman DK, Lebovitz HE. Prostaglandin
 A₁ inhibition of chondrosarcoma growth. Biochem Biophys Res Commun 1974; 60: 1302-8.
- Santoro MG, Philpott GW, Jaffe BM. Inhibition of tumour growth in vivo and in vitro by prostaglandin E. Nature 1976; 263: 777-9.
- Bregman MD, Funk C, Fukushima M. Inhibition of human melanoma growth by prostaglandins A, D, and J analogues. Cancer Res 1986; 46: 2740-4.
- 68. Nakamura O, Maruo K, Ueyama Y, et al. Antineoplastic effect of prostaglandins on human glioma in athymic nude mice (in Japanese). No To Shinkei 1988; 40: 733-7.

H Sasaki and M Fukushima

- Nishimura G. Antitumor activity and cell cycle effects of Δ¹²-prostaglandin J₂ in vivo (in Japanese) Nippon Gan Chiryo Gakkai Shi 1990; 25: 682-9.
- Fukushima M, Kato T, Yamada Y, et al. Inhibition of tumor growth by novel murine eicosanoids, clavulones and punaglandins. Proc Am Ass Cancer Res 1985; 26: 249.
- Kita T, Miyauchi M, Kikuchi Y, et al. Modulation of human lymphocyte response to phytohemagglutinin by antineoplastic prostaglandins (in Japanese). Nippon Gan Gakkai Sokai Kiji 1990; 49: 327.
- 72. Fukushima M. A preclinical study on antitumor prostaglandins. In: *Ninth annual report of 10 year strategy for cancer therapy.* Tokyo: Ministry of Welfare and Health in Japan 1992: 139–44 (in Japanese).
- 73. Hausen HZ. Papilloma viruses as carcinoma viruses. *Adv Viral Oncol* 1989: **8**: 1–26.
- 74. Poiesz BJ. Ruscetti, FW, Gazdar AF, et al. Detection and isolation of type-C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. Proc Natl Acad Sci USA 1980: 77: 7415-9.

- Savona S, Ohri A, Lakkis W, et al. AIDS and malignancy: Experience of an inner city primary care hospital. Proc Am Soc Clin Oncol 1993; 12: 55.
- D'Onofrio C, Amici C, Bonmassar E, et al. The antiproliferative effect of prostaglandin A and J on HTLV-1 transformed cells is associated with induction of a heat-shock protein. Pharmacol Res 1990; 22 (Suppl. 1): 41-2.
- 77. Yamamoto N, Fukushima M, Tsurumi T, et al. Mechanism of inhibition of herpes simplex virus replication by Δ^7 -prostaglandin A_1 and Δ^{12} -prostaglandin J_2 . Biochem Biophys Res Commun 1987; **146**: 1425–31.
- 78. Santoro MG, Jaffe BM, Esteban M. Prostaglandin A inhibits the replication of vesicular stomatitis virus: effect on virus glycoprotein. *J Gen Virol* 1983; **64**: 2797–801.
- Bader TO, Ankel H. Inhibition of primary transcription of vesicular stomatitis virus by prostaglandin A₁. J Gen Virol 1990; 71: 2823-32.

(Received 6 December 1993; accepted 16 December 1993)